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(54) **PREVENTIVES OR REMEDIES FOR DISEASES INDUCED BY HYPOFUNCTION OF NITRIC OXIDE SYNTHASE (NOS)**

VORBEUGENDE ODER HEILENDE MITTEL FÜR KRANKHEITEN, DIE DURCH MANGEL AN STICKOXID-SYNTHASE (NOS) AUSGELÖST SIND

PREPARATIONS SERVANT A LA PREVENTION ET AU TRAITEMENT DE MALADIES PROVOQUEES PAR UNE HYPOFONCTION DE L'OXYDE NITRIQUE SYNTHASE

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(56) References cited:
EP-B1- 0 722 731 **WO-A-95/32203**

- **INOUE, K. ET AL: "Tetrahydrobiopterin abrogates development of DOCA-salt hypertension in rats" JOURNAL OF THE AMERICAN SOCIETY OF NEPHROLOGY, vol. 7, no. 9, September 1996 (1996-09), pages 1534-1535, XP000901298**
- **TIEFENBACHER, C.P. ET AL: "Restoration of endothelium-dependent vasodilation after reperfusion injury by tetrahydrobiopterin" CIRCULATION, vol. 94, no. 6, 15 September 1996 (1996-09-15), pages 1423-1429, XP000901220**
- **PRAST, H. ET AL: "Effects of sepiapterin treatment on tetrahydrobiopterin levels and blood pressure in spontaneously hypertensive rats" BIOLOGY OF NITRIC OXIDE, PROC. INT. MEET., 2ND, vol. 1, 1992, pages 10-12, XP000901316**
- **KUKOR, Z. ET AL: "Calcium-dependent nitric oxide synthesis is potently stimulated by tetrahydrobiopterin in human primordial placenta" PLACENTA, vol. 17, no. 1, January 1996 (1996-01), pages 69-73, XP000901301**
- **TSUTSUI, M.: "Effect of tetrahydrobiopterin on endothelial function in canine middle cerebral arteries" CIRCULATION RESEARCH, vol. 79, no. 2, 1996, pages 336-342, XP000901228**
- **ZVETKOVA, E. ET AL: "Activation and NO secretion pattern of murine macrophage subpopulations after in vivo administration of ranopterin neopterin" COMPTES RENDUS DE L'ACADEMIE BULGARE DES SCIENCES, vol. 49, no. 1, 1996, pages 113-116, XP000901223**

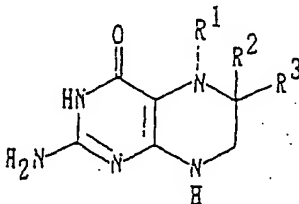
EP 0 908 182 B2

- COSENTINO, F. ET AL: "Tetrahydrobiopterin and endothelial nitric oxide synthase dysfunction in spontaneously hypertensive rats" SUPPLEMENT TO CIRCULATION, vol. 92, no. 8, 1995, page I-565 XP002135728
- PATENT ABSTRACTS OF JAPAN vol. 1995, no. 10, 30 November 1995 (1995-11-30) & JP 07 188233 A (ASAHI BREWERIES LTD.), 25 July 1995 (1995-07-25)
- DATABASE WPI Week 199538 Derwent Publications Ltd., London, GB; AN 1995-290301 XP002135729 & JP 07 188024 A (ASAHI BREWERIES LTD.), 25 July 1995 (1995-07-25)
- BUNE, A.J. ET AL: "Inhibition of tetrahydrobiopterin synthesis reduces nitric oxide production by isolated glomeruli in immune complex glomerulonephritis" EXPERIMENTAL NEUROLOGY, vol. 4, no. 1, 1996, pages 43-47, XP000901378
- AMOAH-APRAKU, B. ET AL: "Guanosine triphosphate cyclohydrolase I regulates nitric oxide synthesis in renal proximal tubules" JOURNAL OF THE AMERICAN SOCIETY OF NEPHROLOGY, vol. 5, no. 8, 1995, pages 1630-1633, XP000901376
- COSENTINO, F. ET AL: "Tetrahydrobiopterin and dysfunction of endothelial nitric oxide synthase in coronary arteries" CIRCULATION, vol. 91, no. 1, 1995, pages 139-144, XP000856340
- HYUN, J. ET AL: "The protective effect of tetrahydrobiopterin on the nitric oxide-mediated inhibition of purified nitric oxide synthase" BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, vol. 206, no. 1, 1995, pages 380-386, XP000891659
- KATUSIC, Z.S. ET AL: "6-methyl-tetrahydropterin selectively inhibits endothelium-dependent contractions in canine basilar artery" FASEB JOURNAL, vol. 8, no. 4-5, 1994, page A826 XP000891677
- DEFILY, D.V. ET AL: "Restoration of endothelium-dependent vasodilation following reperfusion injury by tetrahydrobiopterin" FASEB JOURNAL, vol. 10, no. 3, 1996, page A48 XP000891676
- KOJIMA, S. ET AL: "Neopterin as an endogenous antioxidant" FEBS LETTERS, vol. 304, no. 2,3, June 1992 (1992-06), pages 163-166, XP000892131
- BIOCHEMICAL AND BIOPHYSICAL RESEARCH COMMUNICATIONS, Vol. 215, No. 3, (1995), PEI-FENG CHEN et al., "Cysteine 99 of Endothelial Nitric Oxide Synthase (NOS-III) is Critical for Tetrahydrobiopterin-Dependent NOS-III Stability and Activity", pages 1119-1129.
- JOURNAL OF CARDIOVASCULAR PHARMACOLOGY, Vol. 26, No. 6, (1995), VOLKER VALLON et al., "Interactive Control of Renal Function by alpha2-Adrenergic System and Nitric Oxide: Role of Angiotensin II", pages 916-922.
- SCIENCE. Vol. 272, No. 5262, (1996), GARY H. et al., "Molecular Therapies for Vascular Diseases", pages 689-693.
- LABORATORY INVESTIGATION, Vol. 72, No. 1. (1995), ICHIEI NARITA et al., "Nitric Oxide Mediates Immunologic Injury to Kidney Mesangium in Experimental Glomerulonephritis", pages 17-24.
- ANN. MED., Vol. 27, No. 3, (1995), TIKKANEN IIKKA et al., "Nitric Oxide in Hypertension and Renal Diseases", pages 353-7.
- Scott-Burden, T.: "Regulation of Nitric Oxide Production by Tetrahydrobiopterin", Circulation, vol. 91, 1995, pages 248-250

Description

FIELD OF THE INVENTION

[0001] The present invention relates to a pharmaceutical composition for preventing and/or treating diseases associated with dysfunction of NOS, comprising as an effective ingredient, a compound of the formula (I):



wherein R¹ and R² each represents a hydrogen atom and R³ represents (6R)-L-erythro-CH(OH)CH(OH)CH₃, or a pharmaceutically acceptable salt thereof.

PRIOR ART

[0002] Vascular endothelium has been known to play an important role in vascular tonus or thrombopoiesis, and it was in 1980 that the presence of endothelium-derived relaxing factor (EDRF) was first reported. The entity of EDRF was proved to be nitrogen monoxide (NO) in 1987. NO is produced when L-arginine is oxidized from N^G-hydroxyl-L-arginine into L-citrulline and the reaction is catalyzed by an enzyme named NO synthase (NOS). NOS widely exists in vascular endothelium, nervous systems, kidneys, platelets, cardiac muscles, smooth muscles, etc. and has an important role in control of systemic circulation because the resultant NO has a wide variety of actions. The following diseases with decreased NO production are known: cardiovascular diseases such as hypertension, hyperlipemia, arteriosclerosis, ischemic heart disease, heart failure, thrombosis; respiratory diseases such as asthma, chronic obstructive pulmonary diseases, pulmonary hypertension, ARDS; gastrointestinal diseases such as hepatopathy, liver cirrhosis, gastrointestinal mucosa disorder, hypertrophic pyloric stenosis, pancreatitis; cerebrovascular diseases such as cerebral ischemia, infarction, cerebrovascular failure, senile dementia; renal or urologic diseases such as renal disorder, impotence; gynecological diseases such as toxemia; infectious diseases, immunological disorders, diabetes, burns; or diseases caused by drugs which decrease NO production. The gene for NOS was cloned and structurally analyzed. As a result, the gene for NOS was found to contain a binding site for (6R)-L-erythro-5,6,7,8-tetrahydrobiopterin (hereinafter referred to as "BH4") included in compounds of the formula (I) as active ingredients of the present invention, in addition to those for coenzymes such as calmodulin (CaM), flavin, NADPH. Moreover, BH4 has been suggested to actually be involved in control of the function of NOS.

[0003] On the other hand, it was reported that blood pressure rose in experimental animals which had been intravenously injected with an NOS inhibitor (Sakuma I. et al., Circ. Res. 70: 607-611, 1992). Decreased production of endogenous NO was also reported in animal model of hypertension such as DOCA-salt hypertension rats, SHR-SP rats, Dahl-S rats, Goldblatt hypertension rats.

INOUE, K. ET AL: JOURNAL OF THE AMERICAN SOCIETY OF NEPHROLOGY, vol. 7, No.9, September 1996 (1996-09), pages 1534-1535, discloses that tetrahydrobiopterin abrogates the development of DOCA salt hypertension in rats. Said document is a post-published document of the present inventors.

[0004] Furthermore, PRAST, H. ET AL, BIOLOGY OF NITRIC OXIDE, PROC. INT. MEET., 2ND, vol. 1, 1992, pages 10-12 describes the effects of sepiapterin treatment on tetrahydrobiopterin levels and blood pressure in spontaneously hypertensive rats.

[0005] KUKOR, Z. ET AL, PLACENTA, vol. 17, No. 1, January 1996, pages 69-73, similarly refer to calcium-dependent nitric oxide synthesis being potently stimulated by tetrahydrobiopterin in human primordial placenta. Neither document provides direct evidence showing the pharmacological effects of BH4 on diseases associated with dysfunction of NOS.

[0006] From the article of Amoah-Apraku B. et al. in the Journal of the American Society of Nephrology, vol. 5, No. 8, 1995, pages 1630-1633, it is known that guanosine triphosphate cyclohydrolase regulates nitric oxide synthesis in renal proximal tubules.

[0007] Furthermore, Bune A. J. et al., Experimental Neurology, vol. 4, No. 1, 1996, pages 43 to 47 report that the inhibition of tetrahydrobiopterin synthesis reduces the nitric oxide production by isolated glomeruli in immune complex glomerulonephritis. This disease is associated with increased NOS activity.

[0008] However, the cause for decreased NO production in these models has not been cleared. There are also a

number of reports that NO production is rather enhanced in spontaneous hypertension rats (SHR) which are often used as hypertensive model animals (Dominizak AF et al., Hypertension 25: 1202-1211, 1995). In case of SHR, it may also be assumed that NO is excessively released against hypertensive condition to lower blood pressure but the action of NO is insufficient (Nava E. et al., Circulation 92: 1-347, 1995). Although it has been pointed out that NO may be implicated in the etiology of hypertension, the relationship between the activity of NOS and hypertension has not been fully elucidated yet. The decrease of vasodilative reaction in hypertension may also be caused by the decreased activity of the endothelium-derived hyperdepolarization factor (EDHF) or an increase in the amounts of the endothelium-derived circulating factor (EDCF), both of which have been unknown about their entity.

[0009] Thus, the vasorelaxing action of NO and control of NOS functions by BH4 have been known to some degree. However, the cause for decreased NO production and the relationship between NOS activity and hypertension have not been elucidated, and nothing has been known about antihypertensive action of BH4, as well as the role of BH4 in blood pressure regulation.

[0010] In many industrialized countries, cardiovascular diseases are one of the main causes of death. Various drugs such as antihypertensives, antihyperlipemic agents, diuretics, vasodilators, antiplatelets have been used in the treatment of diseases. However, many of them are designed to improve parameters such as blood pressure or cholesterol level, but still unsatisfactory in the prevention of occurrence, retardation of deterioration or progress and long-term prognosis of these diseases. Recently, on the above-mentioned diseases, by investigating at a molecular level, a therapeutic strategy directed to blood vessels especially endothelial cells has been proposed and as one of the most promising therapies, treatment with an agent which regulates the production of the entity of EDRF, i.e. NO, or an agent which has an antioxidant action has been presented. (Gibbons, G.H., Dzau, V.J., Science. Vol. 272, 689-693, 1996). However, any drug or therapy that satisfies this therapeutic strategy has not been established yet.

[0011] Nitrate agents (nitroglycerin preparations, etc.) have been considered to be effective as exogenous NO donors for treating angina, heart failure or the like among the above diseases, but they show tolerance over long-term use. Namely, thiol groups (SH groups) become depleted during long-term use of nitrate agents, because SH groups are indispensable for NO production from these nitrate agents. Moreover, the balance of other endogenous substances may be disturbed during long-term use, because NO production requires metabolic enzymes and other NO-related materials such as nitrosothiol are produced during the reaction. A new type of NO donors with the properties of β -blockers or K^+ channel openers have also been developed, but they are still within the range of the above-mentioned NO donors. Therefore, improvement of the functions of endothelial cells through regulating endogenous NO function is proposed to be one of the most ideal therapeutic ways for the prevention of occurrence, retardation of deterioration or progress and long-term prognosis of these diseases.

[0012] Spontaneous NO production in vivo or activation of NOS seems to be the best therapeutic method. A substrate for NOS, L-arginine, has been examined for its effects against such diseases as hypertension (Higashi, Y. et al., Hypertension 1995, 25, 898-902), angina (Egashira, K. et al., Circulation 1996, 94, 130-134), heart failure (Rector T.S. et al., Circulation 1996, 93, 2135-2141) or the like.

[0013] However, it was reported that endothelium-dependent vasodilatation reaction and renal NO production were recovered only partially by long-term administration of L-arginine (Hayakawa H., Hirata Y., Omata M. et al., Hypertension 1994; 23: 752-756) in DOCA salt hypertension rats, a well known hypertensive model in which endothelium-dependent vasodilatation reaction by acetylcholine and renal NO production at basal and by the stimulation of endothelin receptors (ET_B) are decreased (Hirata Y., Hayakawa H., Omata M. et al., Circulation 1995; 91: 1229-1235). It was also reported that L-arginine administration resulted in a decrease of blood pressure in hypertension patients similarly to healthy people, but did not bring about an increase in renal bloodflow and the decrease in filtration fractionation and resistance of renal vessels as found in healthy people, and significant decrease in cGMP in blood was observed in hypertension patients (Higashi Y., Ohshima T., Kajiya G. et al., Hypertension 1996; 25: 898-902). These facts suggest that insufficient NO production in renal circulation may neither result from the lack of the substrate L-arginine nor from only the decline of the enzymatic activity of NOS. Accordingly, the participation of L-arginine-NO system in control of renal circulation and the mechanism of decline of endothelium-dependent vasodilatation reaction can not be explained by insufficient utilization or lack of the substrate L-arginine. Therefore, whether the change of the NOS activity or quantity is involved in these phenomenon should be further investigated.

[0014] The purpose of antihypertensive therapy is to prolong the life of patients in a satisfactory condition by preventing cardiovascular complications due to hypertension. For this purpose, blood pressure must be controlled for a long period over life span by antihypertensives in addition to the general therapies such as salt reduction, amelioration of obesity kinesitherapy etc.:

[0015] Major antihypertensives therapeutically used at present include thiazides, β -blockers, Ca-antagonists, angiotensin converting enzyme (ACE) inhibitors and α_1 -blockers. However, none of them satisfy all the following conditions desirable for antihypertensives:

1. Mild action and stable effects;

2. Less side effects at an early stage of use;
3. Desirable influence on circulatory systems and organic functions;
4. No undesirable effects on factors detrimental to cardiovascular systems but rather improvement over long-term use;
5. Better compliance of patients with therapy;
6. No contraindication due to various complications and accidental symptoms;
7. More satisfactory life can be expected as a result of therapy.

[0016] Currently used antihypertensives have the following problems.

[0017] Thiazide-based diuretics are favorably applied when salt limitation is difficult to obey or a tendency of fluid retention is observed, but are not recommended to patients with abnormal glucose tolerance, hyperuricemia, renal dysfunction, hyperlipemia or hypokalemia.

[0018] β -blockers are favorably applied for young or tachycardiac patients, but are not recommended to patients with bronchial asthma, chronic obstructive pulmonary diseases, obstructive lesions in peripheral arteries or Raynaud's symptoms. They are also inappropriate for hypertension patients with diabetes because of the influence on insulin secretion.

[0019] Dihydropyridine derivatives among Ca antagonists have the following side effects, i.e. their vasodilating action may produce facial flush and headache, and their strong hypotensive action may cause hypotension, vertigo and tachycardia or palpitation due to reflex hypersympathicotonus.

[0020] ACE inhibitors have side effects such as orthostatic hypotension, dry cough, vascular edema, hyperkalemia etc. In cases of renal dysfunction, attention should be paid to the dosage and hyperkalemia because they are mainly excreted from kidney. Some of them have been reported to give harmful effects on fetus, and therefore contraindicated for pregnant women.

[0021] α_1 -blockers should be carefully applied so as not to cause side effects such as orthostatic hypotension.

[0022] Thus, no drug is completely satisfactory in terms of side effects, safety during long-term use and improvement in QOL at present, though many antihypertensive agents with various actions exist. In addition, the primary object of antihypertensive therapy consists in preventing the progress of hypertension-induced organic disorders and lowering the incidence of heart failure, renal failure or cerebral apoplexy and the mortality therefrom. Antihypertensive therapy can not prevent an increase in the number of patients with hypertension-induced renal failure, although it was reported that antihypertensive therapy improved the mortality of heart failure and cerebral apoplexy. In acute or chronic glomerulonephritis or the like, hypertension is produced by glomerular disorder. It has been shown that chronic glomerulonephritis with hypertension has a bad prognosis, and hypertension is one of independent prognostic exacerbation factors in chronic glomerulonephritis. However, no therapy for the hypertension with chronic glomerulonephritis has been established.

[0023] Therefore, therapeutic agents providing truly desirable actions are required, and especially the development of agents having a renal protective action are highly demanded.

[0024] The compound of the formula (I) as effective ingredient in the therapeutic agents of the present invention, is a known compound for use in therapeutic agents. For example, refer to Japanese Patent Public Disclosure (KOKAI) Nos. 25325/84, 267781/88 (JP 63 267 781).

SUMMARY OF THE INVENTION

[0025] The object of the present invention is to provide a safe therapeutic agent for diseases associated with dysfunction of NOS which improves circulating and organic functions, retards the progress of complications and improves the quality of life of patients by preventing the decrease of endogenous NO production and regulating the functions of endothelial cells.

[0026] Based on the results of various studies, it is hypothesized that BH4 level might also be lowered in diseases associated with dysfunction of NOS. Thus, BH4 was administered to DOCA-salt hypertension rats and DOCA-salt SHR to test the hypothesis. BH4 level and more surprisingly, NOS expression level were found to be actually lowered in the DOCA-salt hypertension rats. In BH4 administration, decrease of endogenous NO production was prevented and physiologically natural antihypertensive effects were obtained. DOCA-salt SHR also actually exhibited a decline of BH4 level as well as histopathological findings such as necrotizing glomerulitis and necrotizing angiitis. In BH4 administration, antihypertensive effects as well as remarkable improvement effects on said histopathological findings by regulating the functions of endothelial cells were obtained. Thus, the present invention was accomplished on the basis of the finding that BH4 has an action to activate the decreased function of NOS. Accordingly, the present invention is directed to effective therapy with BH4 preparations for diseases associated with dysfunction of NOS.

BRIEF DESCRIPTION OF THE DRAWINGS

[0027]

FIG. 1 shows the time course of blood pressure change in a control group (-○-), DOCA group (-Δ-) and a group of DOCA-salt hypertension rats orally administered BH4 (DOCA + BH4 group, -□-).

FIG. 2 shows the time course of change of NO₂/NO₃ content in urine in a control group (-○-), DOCA group (-Δ-) and a group of DOCA-salt hypertension rats orally administered BH4 (DOCA + BH4 group, -□-).

FIG. 3 shows the time course of change of BP content in urine in a control group (-○-), DOCA group (-Δ-) and a group of DOCA-salt hypertension rats orally administered BH4 (DOCA + BH4 group, -□-).

FIG. 4 shows the results of measurement of the expression of E-NOS mRNA in renal tissues.

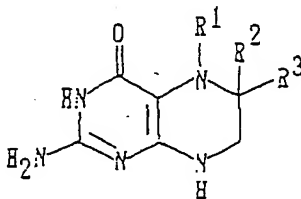
FIG. 5 shows the time course of blood pressure change in a control group (-●-), DOCA group (-Δ-) and a group of DOCA-salt SHR rats orally administered BH4 (DOCA + BH4 group, -□-).

FIG. 6 shows the time course of change of BP content in urine in a control group (-●-), DOCA group (-Δ-) and a group of DOCA-salt SHR rats orally administered BH4 (DOCA + BH4 group, -□-).

FIG. 7 shows the incidences of histopathological findings in renal tissue samples on necrotizing glomerulitis and necrotizing angitis in a control group, DOCA group and DOCA + BH4 group.

DETAILED DESCRIPTION OF THE INVENTION

[0028] The present invention relates to a pharmaceutical composition for preventing and/or treating diseases associated with dysfunction of NOS, comprising as an effective ingredient, a compound of the formula (I) :



wherein R¹ and R² each represents a hydrogen atom and R³ represents (6R)-L-erythro -CH(OH)CH(OH)CH₃, or a pharmaceutically acceptable salt thereof.

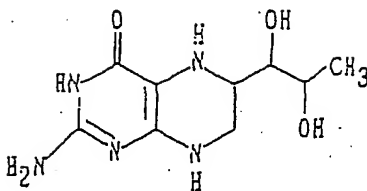
[0029] As used herein, dysfunction of NOS means that the expression of NOS, which widely occurs in vascular endothelium, nervous systems, kidney, platelets, myocardium, smooth muscles or other organs, has been reduced for some reason or the activity of NOS is not exhibited by dysfunction of these cells even if it is expressed. A typical example of dysfunction of NOS is a decrease in the endogenous NO level. Diseases associated with dysfunction of NOS include those induced, deteriorated or hindered from cure by dysfunction of NOS, such as hypertension, hyperlipemia, arteriosclerosis, coronary vasospasm, ischemic heart disease, heart failure, thrombosis, pulmonary hypertension, cerebrovascular failure, cerebral vasospasm, glomerulonephritis, chronic renal failure, diabetes, postoperative restenosis, achalasia, portal hypertension, hepatic disorder, gastrointestinal mucosa disorder, hypertrophic pyloric stenosis, enteritis, impotence, chorioretinopathy, etc.

[0030] When administered to patients with these diseases, BH4 can prevent or treat these diseases by normalizing the functions of NOS through the stimulating effect on NOS production in vivo or through restoring the decreased function of endothelial cells.

[0031] Accordingly, the present invention is applied to treat or prevent diseases that can be treated by the stimulatory action of BH4 to activate the function of NOS.

[0032] The compound of the formula (I) as effective ingredient of the present invention is the following one and pharmaceutically acceptable salts thereof:

(6R)-L-erythro-5,6,7,8-tetrahydrobiopterin (BH4)



[0033] The compound of the formula (I) used as effective ingredient in the present invention is a known compound. For example, refer to Japanese Patent Public Disclosure (KOKAI) Nos. 25323/84, 267781/88 (JP 63 267 781). The compound may be used as appropriate salts with pharmacologically non-toxic acids, including mineral acids such as hydrochloric acid, phosphoric acid, sulfuric acid, boric acid; and organic acids such as acetic acid, formic acid, maleic acid, fumaric acid, mesylic acid.

[0034] A pharmaceutical composition of the present invention is effective against the above mentioned diseases associated with dysfunctional Nos. For example, it is effective against hypertension including not only essential hypertension but also malignant hypertension with a necrosis of arteriolar walls in the kidney and retina occurring in a short period, and renal hypertension accompanied by renal disorder. Namely, it is effective against all kinds of hypertension such as essential hypertension, renal hypertension, renovascular hypertension, pregnancy-induced hypertension, senile hypertension and adrenal hypertension, as long as they are associated with dysfunctional Nos.

[0035] A pharmaceutical composition of the present invention is prepared by formulating a compound of the formula (I) with generally used carriers by conventional procedures into a dosage form suitable for oral, rectal or parenteral administration (including administration into vein and cerebrospinal fluid).

[0036] The carrier used for these pharmaceutical formulations generally include excipients, binders, disintegrators, etc. depending on the dosage form chosen.

[0037] Typical examples of excipients include starch, lactose, sucrose, glucose, mannitol and cellulose, and examples of binders include polyvinylpyrrolidone, starch, sucrose, hydroxypropyl cellulose, Arabic gum. Examples of disintegrators include starch, agar, gelatin powder, cellulose, CMC, but any other conventional excipients, binders and disintegrators may also be used.

[0038] In addition to such carriers, pharmaceutical composition of the present invention may also contain antioxidants for stabilizing effective ingredients. Antioxidants can be appropriately selected from those conventionally used for pharmaceutical preparations, such as ascorbic acid, N-acetylcysteine, L-cysteine, dl- α -tocopherol, natural tocopherol etc. They are used in an amount that stabilizes the active ingredient and generally, the ratio of an antioxidant between 0.2 and 2.0 parts by weight to 1 part of the active ingredient.

[0039] Formulations of the present invention suitable for oral administration may be provided in the form of tablets, sublingual tablets, capsules, powders, granules or fine granules, or suspensions in a non-aqueous liquid such as syrups, emulsions or draft (pro re nata preparation) that contain the prescribed amount of the active ingredient.

[0040] For example, granules are prepared by homogeneously mixing the active ingredient with one or more auxiliary ingredients such as carriers and antioxidants as mentioned above, followed by granulation and sieving to uniform grain size. Tablets can be prepared by compressing or molding the active ingredient optionally together with one or more auxiliary ingredients. Capsules are prepared by filling powder or granules of active ingredient optionally mixed homogeneously with auxiliary ingredient (one or more) into appropriate capsules using a capsule filling machine or the like. Formulations for rectal administration can be provided as suppositories using conventional carriers such as cacao butter. Parenteral formulations can be provided as dry solids of active ingredient sealed in a nitrogen-filled sterilized containers. Such dry solid preparations can be administered to patients after dispersing or dissolving them into a determined amount of sterilized water just prior to administration.

[0041] In manufacturing these formulations, antioxidants as mentioned above may preferably added to a mixture of effective ingredient and conventional carriers, and optionally one or more auxiliary ingredients selected from buffers, flavors, surfactants, viscosants, lubricants, etc. can also be added, if needed.

[0042] The dosage of active ingredients i.e. the compound of the formula (I) may vary with the administration route, the symptom to be treated and the patient condition, and the final determination of the dosage should be made by an attendant physician.

[0043] For example, an appropriate dosage for treating hypertension ranges from 0.1 to 50 mg/kg (b.w.) / day, and representative optimal dosage is 0.5 to 10 mg/kg (b.w.) / day.

[0044] A desired dosage of said active ingredients may be administered once a day or in divided doses of two to four times a day at appropriate interval.

[0045] The active ingredient may be administered alone or in combination with pharmaceutical formulations containing other active ingredients suitable for the disease under treatment to facilitate control of the dosage, for example.

[0046] In addition to the compound of the formula (I) as effective ingredient, formulations of the present invention may contain at least one auxiliary effective ingredient selected from the substrates or coenzyme or cofactor for NOS such as L-arginine flavins, for example, FAD, FMN, etc., or calcium. More excellent therapeutic effects can be expected when the compound of the formula (I) is mixed with these effective ingredients than when used alone. The proportion of each of said auxiliary effective ingredients in formulations of the present invention is not specifically limited. For example, the weight ratio of at least one selected from L-arginine, flavins and calcium to 1 part of the compounds of the formula (I) may be within the range from 0.1 to 10, preferably 0.5 to 2.

[0047] For example, an appropriate dosage of such mixed formulations for treating hypertension ranges from 0.1 to 50 mg/kg (b.w.) / day, preferably 0.5 to 10 mg/kg (b.w.) / day, in terms of the total amount of active ingredients.

[0048] A physician may appropriately choose formulations containing the compound of the formula (I) alone or in combination with other active ingredients, depending on the age, condition or other factors of the patient.

[0049] The active ingredient used in the present invention is (6R)-L-erythro-5,6,7,8-tetrahydrobiopterin (BH4) and salts thereof. Needless to say, BH4, naturally occurring component is most preferable. BH4 dihydrochloride has little toxicity to rats judging from the fact that an acute toxicity was more than 2 g/kg (b.w.). An optically inactive (6R,S)-5,6,7,8-tetrahydrobiopterin is also only slightly toxic as reported in Japanese Patent Public Disclosure No. 25323/84 for the treatment of Parkinson's disease, so that it can also be used according to the present invention.

[0050] The following examples further explain the present invention in detail, but the present invention is not limited to these examples.

EXAMPLES

Example 1: Granules and fine granules

[0051] To 1 part (by weight) of polyvinylpyrrolidone (Kollidon 30) dissolved in sterilized purified water were added 10 parts of ascorbic acid and 5 parts of L-cysteine hydrochloride to give a homogeneous solution, and then 10 parts of BH4 dihydrochloride were added to prepare a homogeneous solution.

[0052] This solution was added to 59 parts of an excipient (mannitol or lactose) and 15 parts of a disintegrant [corn starch or hydroxypropylcellulose (LH-22)], and the mixture was kneaded, granulated, dried, then sieved.

Example 2: Tablets

[0053] The homogeneous solution of an active ingredient prepared in Example 1 was mixed with 58 parts of lactose and 15 parts of microcrystalline cellulose, then with 1 part of magnesium stearate and tableted.

Example 3: Capsules

[0054] The dosage form prepared in Example 1 filled into capsules, wherein the formulation include 0.2% of magnesium stearate as a lubricant.

Example 4: Injection

[0055]

BH4 dihydrochloride	1.5 g
Ascorbic acid	1.5 g
L-cysteine hydrochloride	0.5g
Mannitol	6.5 g

[0056] The above ingredients were dissolved into sterile purified water to make 100 ml and then sterilized, and 1 ml or 2 ml aliquot each was dispensed into a vial or ampule, then lyophilized and sealed.

Example 5: Injection

[0057] 2.0 g of BH4 dihydrochloride was dissolved in sterile purified water to make 100 ml under an anaerobic atmosphere and then sterilized and sealed in the same way as in Example 4.

Example 6: Suppositories**[0058]**

5

BH4 dihydrochloride	150 parts
Ascorbic acid	150 parts
L-cysteine hydrochloride	50 parts

10 **[0059]** The above ingredients were homogeneously ground and dispersed into 9950 parts of cacao butter.

Example 7: Granules**[0060]**

15

BH4 dihydrochloride	5 parts
Ascorbic acid	5 parts
L-cysteine hydrochloride	2 parts

20

[0061] The above ingredients were used to prepare a homogeneous solution.

[0062] This solution was added to a homogeneous mixture of 55 parts of mannitol, 1 part of polyvinylpyrrolidone, 14 parts of hydroxypropylcellulose and 5 parts of L-arginine or calcium, and the mixture was kneaded, granulated, dried, then sieved.

25

Example 8: Granules**[0063]**

30

BH4 dihydrochloride	5 parts
Ascorbic acid	5 parts
L-cysteine hydrochloride	5 parts
Mannitol	52 parts
Polyvinylpyrrolidone (Kollidon 30)	1 part
Hydroxypropylcellulose (LH-22)	12 parts
L-arginine or calcium	10 parts

35

40 **[0064]** The above ingredients were granulated and sieved in the same way as in Example 7.

Example 9: Granules**[0065]**

45

BH4 dihydrochloride	5 parts
Ascorbic acid	5 parts
L-cysteine hydrochloride	2 parts

50

[0066] The above ingredients were used to prepare a homogeneous solution.

[0067] This solution was added to a homogeneous mixture of 10 parts of L-arginine or calcium, 50 parts of mannitol, 1 part of polyvinylpyrrolidone (Kollidon 30) and 9 parts of hydroxypropylcellulose (LH-22), and the mixture was kneaded, granulated, dried, then sieved.

55

Example 10: Preparation of DOCA-salt hypertension rats

[0068] One kidney was removed from each of 8-week-old Sprague-Dawley rats (supplied from Charles River) except

for a control group. Then, the animals were divided into the following three experimental groups.

- 1) A group given drinking water only (control group); sample size $n = 3$;
- 2) A group of DOCA-salt hypertension rats prepared by giving 30 mg/kg b.w. of deoxycorticosterone acetate (DOCA) subcutaneously once a week and 1%-saline as drinking water from one week after nephrectomy (DOCA group); $n = 6$;
- 3) A group of DOCA-salt hypertension rats administered BH4 (10 mg/kg b.w. / day) orally (DOCA + BH4 group); $n = 5$.

[0069] Urine was collected for 24 hours using a metabolic cage at the start of experiment and after 2, 4 and 5 weeks (at the termination of the observation period). At the termination of the observation period, blood sample was collected from abdominal aorta under anesthesia with Phenobarbital. The kidney was isolated after perfusing with physiological saline, and examined histologically and immunohistologically.

Measurement of blood pressure

[0070] Blood pressure was measured by the tail-cuff method at the start of experiment and 2, 4 and 5 weeks.

Measurement of NO_2/NO_3 in urine

[0071] The measurement was carried out using Nitrate-Nitrite assay kit, (kit No. 780001, Cyman Chemical Company) according to the Griess method. The values of NO_2/NO_3 in urine means the activity of NOS, namely, higher the value, higher the activity.

Measurement of BP in urine

[0072] Biopterin (BP), a metabolite of BH4, was measured by a HPLC according to the procedure of Fukushima and Nixon (Anal. Biochem. 102: 176-188, 1980). The effects of BH4 administration are evaluated on the basis of change of BP level in urine.

Immunostaining of renal tissue NOS

[0073] The expression of endothelial NOS (e-NOS) and brain NOS (b-NOS) in renal tissues was examined in the sections of the isolated frozen renal tissues by immunostaining with anti-e-NOS and anti-b-NOS antibodies (Affinity Bioreagents), respectively.

Measurement of NOS mRNA expression

a) Extraction of whole RNA from renal tissues

[0074] Extraction of RNA from renal tissues was performed by the guanidium/cesium chloride extraction method.

[0075] Namely, a small amount of tissues collected from renal cortex of the isolated kidney was added immediately to a lysis solution 4M guanidinium thiocyanate and homogenized by a glass/Teflon homogenizer. This lysate was overlaid on 5.7M cesium chloride and centrifuged at 80,000 rpm for 2 hours, then the precipitate was dissolved in 0.1% diethyl pyrocarbonate (DEP) solution. This solution was precipitated with 2.5M ammonium acetate and ethanol at -20°C for one hour and centrifuged at 10,000 rpm at 4°C for 20 minutes to give a precipitate. The precipitate was dissolved in 0.1% DEP solution at the concentration of about $1 \mu\text{g}/\mu\text{l}$ and stored at -20°C before use.

b) Reverse transcriptase (RT) reaction

[0076] cDNA was synthesized from whole RNA by using MoMLV reverse transcriptase.

[0077] In summary, a mixture of whole RNA, primers, buffer and H_2O was heated at 95°C for 2 minutes and then ice-cooled. Then, the mixture was incubated at 37°C for 30 minutes to anneal the primers to RNA. Then, a solution containing, 0.2M dithiothreitol (DTT), and 5 mM deoxyribonucleotide (dNTP), RNAasin, and MoMLV reverse transcriptase were added to the mixture. The resulted mixture was heated at 90°C for 5 minutes and the reaction was stopped by ice-cooling.

c) PCR

[0078] PCR was performed using cDNA synthesized by the RT reaction as a template.

[0079] Specifically, the RT reaction solution was added to a mixture of 5'- and 3'-primers for each of e-NOS to be

detected and for malate dehydrogenase (MDH) used as an internal standard, PCR buffer, 2.5 mM dNTP mixed solution and Taq DNA polymerase. The reaction mixture was overlaid with mineral oil and centrifuged, after which the reaction was started in an automatic PCR apparatus. PCR products were obtained by running 30 cycles wherein each cycle consists of 94°C for 30 seconds, 50°C for 30 seconds and 72°C for 30 seconds. The primers used for the detection of e-NOS and MDH were selected based on the report by Ujiié, K. et al., Am. J. Physiol. 267 (Renal Fluid Electrolyte Physiol. 36): F296-F302. 1994. Specifically, the base sequences of the primers used for the detection of e-NOS were TACGGAGCAGCAAATCCAC (SEQ ID NO: 1) for 5' primer and CAGGCTGCAGTCCTTTGATC (SEQ ID NO: 2) for 3' primer. The base sequences of the primers used for the detection of MDH were CAAGAAGCATGGCGTATACAACCC (SEQ ID NO: 3) for 5' primer and TTTCAGCTCAGGGATGGCCTCG (SEQ ID NO: 4) for 3' primer.

d) Measurement of NOS mRNA expression by polyacrylamide gel electrophoresis and Southern hybridization

[0080] PCR products were electrophoresed on 5 % polyacrylamide gel and NOS mRNA expression was measured by Southern hybridization.

[0081] Specifically, the PCR reaction solution was electrophoresed on 5% polyacrylamide and then, the gel was transferred to membrane filters. Hybridization was performed on the membrane filters using probes labeled with $\gamma^{32}\text{P}$ -ATP at the 5' prime for the detection of e-NOS and MDH (Ujiié, K. et al., 1994, supra.). The base sequence of the probe was CTGGAACAATTTCCATCCG (SEQ ID NO: 5) for the detection of e-NOS and TTTGTCTTCTCCCTGGTGA (SEQ ID NO: 6) for the detection of MDH. Then, autoradiography was run at -70°C for several hours and the exposed autoradiograms were analyzed by a densitometer. The results are shown in Fig. 4.

[0082] On the basis of the results of autoradiography, the amount of e-NOS PCR products was normalized against the amount of housekeeping gene MDH PCR products to semiquantify e-NOS gene expression.

Statistics

[0083] The measured values were expressed as means \pm standard error and evaluated statistically using two-way ANOVA test. In the following analyses, the significant level sets $p < 0.05$.

[0084] Change of blood pressure is shown in Table 1 and Fig. 1. DOCA group developed hypertension at week 4 after the start of the observation. The rise in blood pressure in DOCA group was significant as compared with the other two groups. However, DOCA + BH4 group stayed at normal blood pressures similarly to the control group until the termination of the observation period.

Table 1

Blood pressure (mmHg)				
	Start	Week2	Week4	Week 5
Control group	101.38 (± 7.12)	102.85 (± 11.07)	91.77 (± 7.88)	111.70 (± 10.10)
DOCA group	107.03 (± 2.04)	137.53 (± 4.63)	153.53 (± 7.71)	150.68 (± 3.35)
DOCA + BH4 group	105.72 (± 5.63)	120.54 (± 6.57)	111.91 (± 7.52)	107.78 (± 7.93)

[0085] Change of NO_2/NO_3 in urine is shown in Table 2 and Fig. 2. In DOCA group, the value of NO_2/NO_3 in urine decreased with time and fell below the measurement sensitivity at the termination of the observation period. However, DOCA + BH4 group showed an increase in the value. The decrease in DOCA group was significant as compared with the other two groups, and no significant difference was found between the control group and DOCA + BH4 group.

Table 2

NO_2/NO_3 in urine (mmol / day)				
	Start	Week 2	Week 4	Week 5
Control group	2.91 ± 0.09		1.51 ± 0.79	4.54 ± 0.32
DOCA group	3.11 ± 1.09		0.34 ± 0.34	below detection limit
DOCA + BH4 group	2.22 ± 0.70		2.00 ± 0.55	5.95 ± 1.90

Change of BP in urine is shown in Table 3 and Fig. 3. In DOCA group, the value of BP decreased with time. However, DOCA + BH4 group showed an increase in the value of BP with time. The decrease in DOCA group was significant as compared with the other two groups, and no significant difference was found between the control group and DOCA + BH4 group.

Table 3

BP in urine (nmol / day)				
	Start	Week 2	Week 4	Week 5
Control group	26.76±3.01	21.71±0.61	27.79± 2.03	34.96±0.96
DOCA group	23.77±0.77	19.49±2.52	15.93± 3.46	19.25±1.96
DOCA+BH4 group	21.87±0.60	35.58±3.74	52.60±13.13	34.26±4.08

[0086] As shown in Table 4, the body weight increased with time in all the groups and no significant difference was found among the three groups.

Table 4

Body weight (g)				
	Start	Week 2	Week 4	Week 5
Control group	390.67±18.49	433.33±21.49	456.00±21.63	458.00±20.43
DOCA group	376.00± 3.69	399.67±19.44	430.33±20.08	435.67±23.34
DOCA+BH4 group	383.40±10.83	386.40±12.84	431.20±13.31	446.80±15.81

[0087] The results of immunostaining showed that the expression of e-NOS in small vessels and the expression of b-NOS in macula densa both decreased in DOCA group while these expressions in DOCA + BH4 group were comparable to the control group or slightly enhanced.

(Results of measurement of NOS mRNA expression)

[0088] As shown in Fig. 4, the results of measurement revealed that the expression of e-NOS mRNA was significantly enhanced in DOCA + BH4 group as compared with DOCA group.

[0089] As described above, DOCA-salt hypertension rats exhibited a rise in blood pressure, a decrease in NO₂/NO₃ in urine and a decrease in NOS expression in renal tissues, indicating that dysfunction of vasoendothelial cells occurred in this model rats. A decrease in BP in urine was also observed in this model rats. On the other hand, oral administration of BH4 suppressed the rise in blood pressure and led to an increase of NO₂/NO₃ in urine and an increase of NOS expression in renal tissues, indicating that BH4 is useful for the treatment of diseases caused by decreased endogenous NO production through activating the function of NOS (through regulation of NOS level/ NOS activity).

Example 11: Preparation of DOCA-salt SHR

[0090] One kidney was removed from 8-week old SHRs (supplied from Charles River) except for a control group. Then, the animals were divided into the following three experimental groups.

- 1) A group given drinking water only (control group); sample size n = 8;
- 2) A group of DOCA-salt SHRs prepared by giving 30 mg/kg b.w. of deoxycorticosterone acetate (DOCA) subcutaneously once a week and 1% saline as drinking water from one week after nephrectomy (DOCA group); n = 4;
- 3) A group of DOCA-salt SHRs administered BH4 (10 mg/kg b.w. / day) orally (DOCA + BH4 group); n = 6.

[0091] Urine was collected for 24 hours using a metabolic cage at the start of experiment and after 2, 3, 4, 5 and 6 (at the termination of the observation period) weeks. At the termination of the observation period, blood sample was collected from abdominal aorta under anesthesia with Phenobarbital. The kidney was isolated after perfusing with physiological saline, and examined histologically.

Measurement of blood pressure

[0092] Blood pressure was measured by the tail-cuff method at the start of experiments and after 2, 3, 4, 5 and 6 weeks.

Measurement of BP in urine

[0093] Biopterin (BP), a metabolite of BH4, was measured by a HPLC according to the procedure of Fukushima and Nixon. The effects of BH4 administration are evaluated on the basis of change of BP level in urine.

Histopathological findings

[0094] After collecting urine and measuring blood pressure at 6 weeks, the animals was incised under anesthesia, exsanguinated from abdominal aorta and perfused with physiological saline via this site. The isolated kidney was fixed in a 10 % formaldehyde solution, embedded in paraffin, then sliced to 4 μ m in thickness by a microtome. Samples were stained with hematoxylin/eosin (HE) and periodic acid-Shiff. The incidences of histopathological findings on necrotizing glomerulitis and necrotizing arteriolitis were compared among the groups.

Statistics

[0095] The measured values were expressed as means \pm standard error and evaluated statistically using two-way ANOVA test. In the following analyses, the significant level sets $p < 0.05$.

[0096] Change of blood pressure is shown in Table 5 and Fig. 5. DOCA group developed remarkable hypertension at week 3 after the start of the observation. The rise in blood pressure in DOCA group was significant as compared with the other two groups. However, DOCA + BH4 group showed the same blood pressures level as that of the control group until the termination of the observation period.

Table 5

Blood pressure (mmHg)						
	Start	Week 2	Week 3	Week 4	Week 5	Week 6
Control group	129.44 (± 2.20)	119.44 (± 2.97)	143.83 (± 0.62)	151.20 (± 0.90)	158.01 (± 1.63)	161.91 (± 4.20)
DOCA group	132.00 (± 5.18)	132.88 (± 3.88)	162.28 (± 1.87)	169.83 (± 2.57)	182.98 (± 2.04)	178.10 (± 9.85)
DOCA + BH4 group	130.22 (± 3.75)	133.60 (± 2.05)	143.63 (± 0.85)	150.90 (± 0.94)	159.30 (± 2.84)	162.95 (± 1.52)

[0097] Change of BP in urine is shown in Table 6 and Fig. 6. In DOCA group, the value of BP decreased with time. However, DOCA + BH4 group showed an increase in the value of BP with time. The decrease in DOCA group was significant as compared with the other two groups, and no significant difference was found between the control group and DOCA + BH4 group.

Table 6

BP in urine (nmol / day)			
	Start	Week 4	Week 6
Control group	22.29 \pm 1.53	25.13 \pm 1.23	43.07 \pm 7.24
DOCA group	17.87 \pm 1.65	3.18 \pm 2.14	16.33 \pm 5.77
DOCA + BH4 group	24.26 \pm 1.58	45.21 \pm 10.31	71.78 \pm 32.08

[0098] The incidences of histopathological findings in renal tissue samples are shown in Table 7 and Fig. 7. DOCA group showed significantly higher incidences of necrotizing glomerulitis and necrotizing angiitis as compared with the other two groups.

Table 7

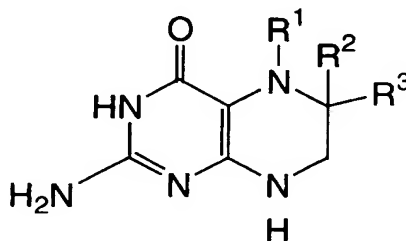
Percentage (%) of incidences of histopathological findings in renal tissues		
	Necrotizing glomerulitis	Necrotizing angiitis
Control group	0.06±0.06	0.00±0.00
DOCA group	10.78±0.77	9.03±1.65
DOCA + BH4 group	2.47±1.07	3.34±0.94

[0099] As described above, DOCA-salt SHR exhibited a rise in blood pressure and histopathological findings of necrotizing glomerulitis or necrotizing angiitis. A decrease in BP in urine was also observed in this model rats. On the other hand, oral administration of BH4 suppressed the rise in blood pressure and remarkably improved histopathological findings, indicating that BH4 is useful for the treatment of diseases associated with dysfunction of endothelial cells through activating the function of NOS.

[0100] As has been explained, the present invention provides a pharmaceutical composition agents that effectively prevents and/or ameliorates diseases associated with dysfunction of NOS. In addition, the active ingredient of the therapeutic agent of the present invention have no adverse effects or the like over long-term use because it is a substance that inherently exists in vivo.

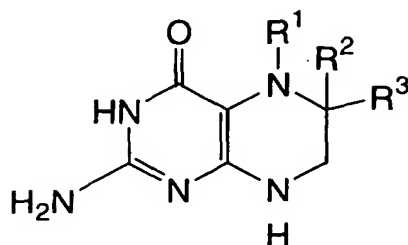
Claims

1. Use of a compound of the formula (I):



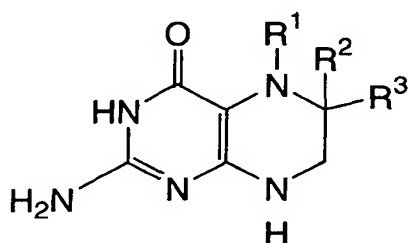
wherein R¹ and R² each represents a hydrogen atom, and R³ represents (6R)-L-erythro-CH(OH)CH(OH)CH₃, or a pharmaceutically acceptable salt thereof for the preparation of a medicament for preventing or treating diseases associated with dysfunction of NOS.

2. Use according to claim 1, wherein the disease associated with dysfunction of NOS is a disease accompanied by decreased endogenous NO production.
3. Use according to claim 1, wherein the disease associated with dysfunction of NOS is a disease accompanied by dysfunction of endothelial cells.
4. Use according to claim 1, wherein the disease associated with dysfunction of NOS is associated with a decrease in NOS expression.
5. Use of a compound of the formula (I):



wherein R^1 and R^2 each represents a hydrogen atom, and R^3 represents (6R)-L-erythro-CH(OH)CH(OH)CH₃, or a pharmaceutically acceptable salt thereof for the preparation of a prophylactic or therapeutic agent for hypertension associated with dysfunction of NOS.

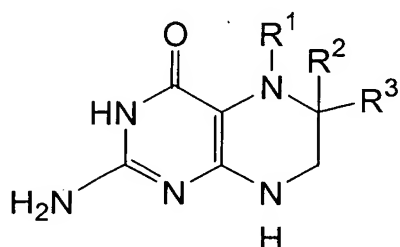
6. Use of a compound of the formula (I):



wherein R^1 and R^2 each represents a hydrogen atom, and R^3 represents (6R)-L-erythro-CH(OH)CH(OH)CH₃, or a pharmaceutically acceptable salt thereof for the preparation of a prophylactic or therapeutic agent for renal disorders associated with dysfunction of NOS.

Patentansprüche

1. Verwendung einer Verbindung der Formel (I):



worin R^1 und R^2 jeweils ein Wasserstoffatom darstellen und R^3 (6R)-L-Erythro-CH(OH)CH(OH)CH₃ darstellt, oder eines pharmazeutisch annehmbaren Salzes davon, zur Herstellung eines Medikaments zur Verhinderung oder Behandlung von Erkrankungen, die mit einer Dysfunktion von NOS assoziiert sind.

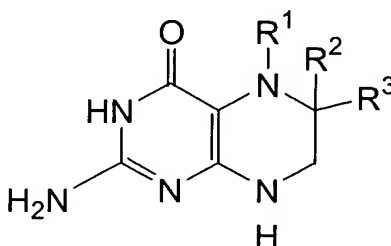
2. Verwendung gemäß Anspruch 1, wobei die Erkrankung, die mit einer Dysfunktion von NOS assoziiert ist, eine Erkrankung ist, die von einer verminderten endogenen NO-Produktion begleitet wird.

3. Verwendung gemäß Anspruch 1, wobei die Erkrankung, die mit einer Dysfunktion von NOS assoziiert ist, eine

Erkrankung ist, die von einer Dysfunktion der Endothelzellen begleitet wird.

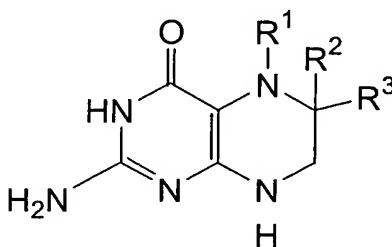
4. Verwendung gemäß Anspruch 1, wobei die Erkrankung, die mit einer Dysfunktion von NOS assoziiert ist, mit einer Abnahme der NOS-Expression assoziiert ist.

5. Verwendung einer Verbindung der Formel (I):



worin R^1 und R^2 jeweils ein Wasserstoffatom darstellen und R^3 (6R)-L-Erythro-CH(OH)CH(OH)CH₃ darstellt, oder eines pharmazeutisch annehmbaren Salzes davon, zur Herstellung eines prophylaktischen oder therapeutischen Mittels für Bluthochdruck, der mit einer Dysfunktion von NOS assoziiert ist.

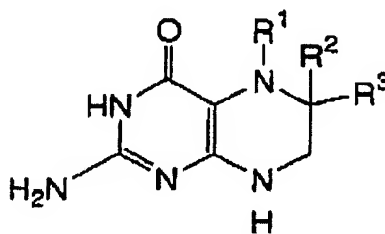
6. Verwendung einer Verbindung der Formel (1):



worin R^1 und R^2 jeweils ein Wasserstoffatom darstellen und R^3 (6R)-L-Erythro-CH(OH)CH(OH)CH₃ darstellt, oder eines pharmazeutisch annehmbaren Salzes davon, zur Herstellung eines prophylaktischen oder therapeutischen Mittels für Nierenstörungen, die mit einer Dysfunktion von NOS assoziiert ist.

Revendications

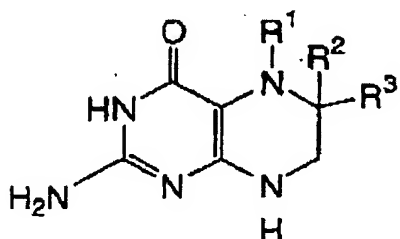
1. Utilisation d'un composé de la formule (I) :



dans laquelle R^1 et R^2 représentent chacun un atome d'hydrogène, et R^3 représente un (6R)-L-érythro-CH(OH)CH

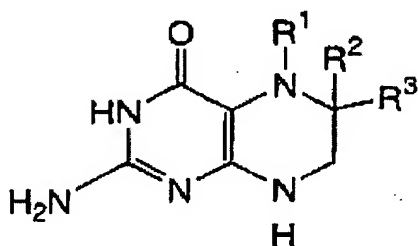
(OH)CH₃, ou un sel pharmaceutiquement acceptable de celui-ci pour la préparation d'un médicament destiné à prévenir ou à traiter les maladies associées au dysfonctionnement du NOS.

2. Utilisation selon la revendication 1, dans laquelle la maladie associée au dysfonctionnement du NOS est une maladie accompagnée d'une production décreue de NO endogène.
3. Utilisation selon la revendication 1, dans laquelle la maladie associée au dysfonctionnement du NOS est une maladie accompagnée d'un dysfonctionnement des cellules endothéliales.
4. Utilisation selon la revendication 1, dans laquelle la maladie associée au dysfonctionnement du NOS est associée à une diminution dans l'expression des NOS.
5. Utilisation d'un composé de la formule (I) :



dans laquelle R¹ et R² représentent chacun un atome d'hydrogène, et R³ représente un (6R)-L-érythro-CH(OH)CH(OH)CH₃, ou un sel pharmaceutiquement acceptable de celui-ci pour la préparation d'un agent prophylactique ou thérapeutique pour l'hypertension associée au dysfonctionnement du NOS.

6. Utilisation d'un composé de la formule (I) :



dans laquelle R¹ et R² représentent chacun un atome d'hydrogène, et R³ représente un (6R)-L-érythro-CH(OH)CH(OH)CH₃, ou un sel pharmaceutiquement acceptable de celui-ci pour la préparation d'un agent prophylactique ou thérapeutique pour les troubles rénaux associés au dysfonctionnement du NOS.

Fig. 1

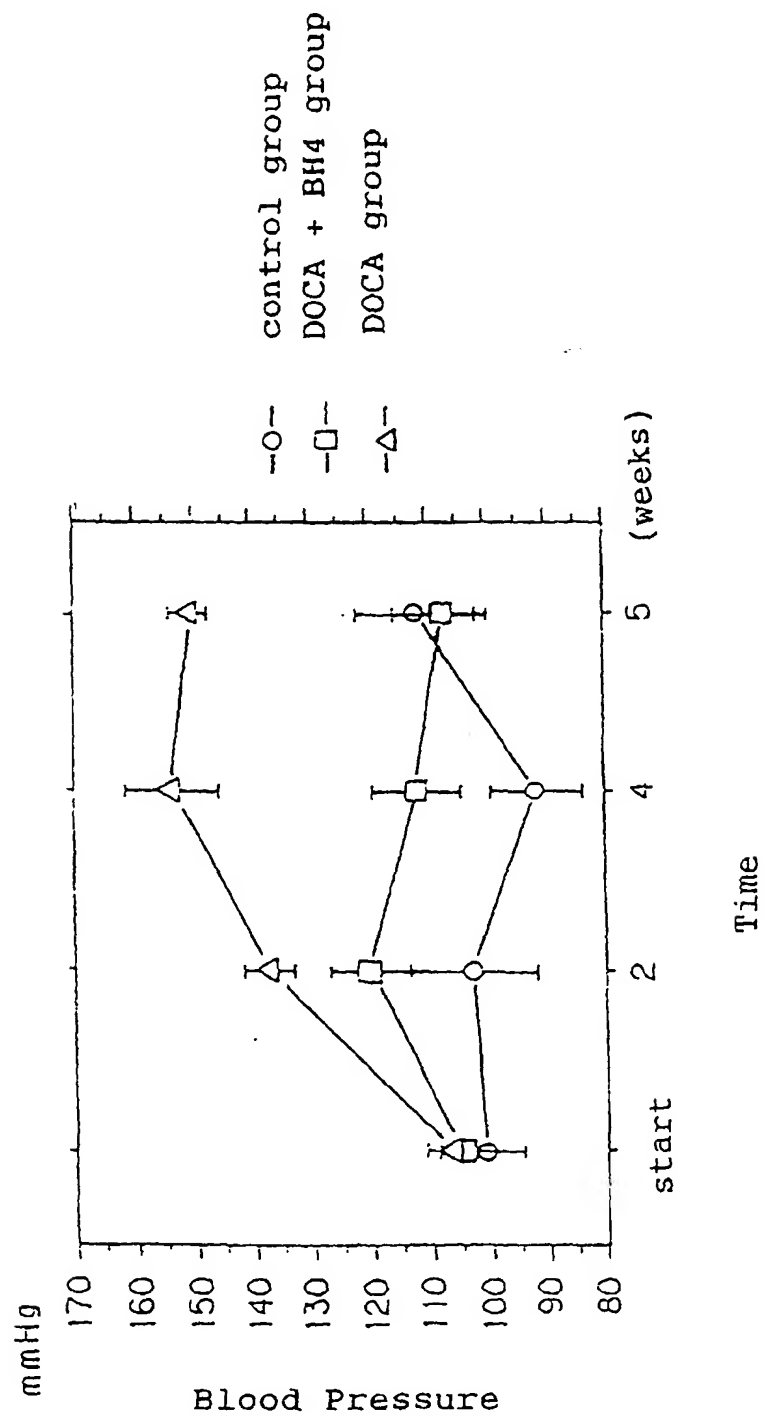


Fig. 2

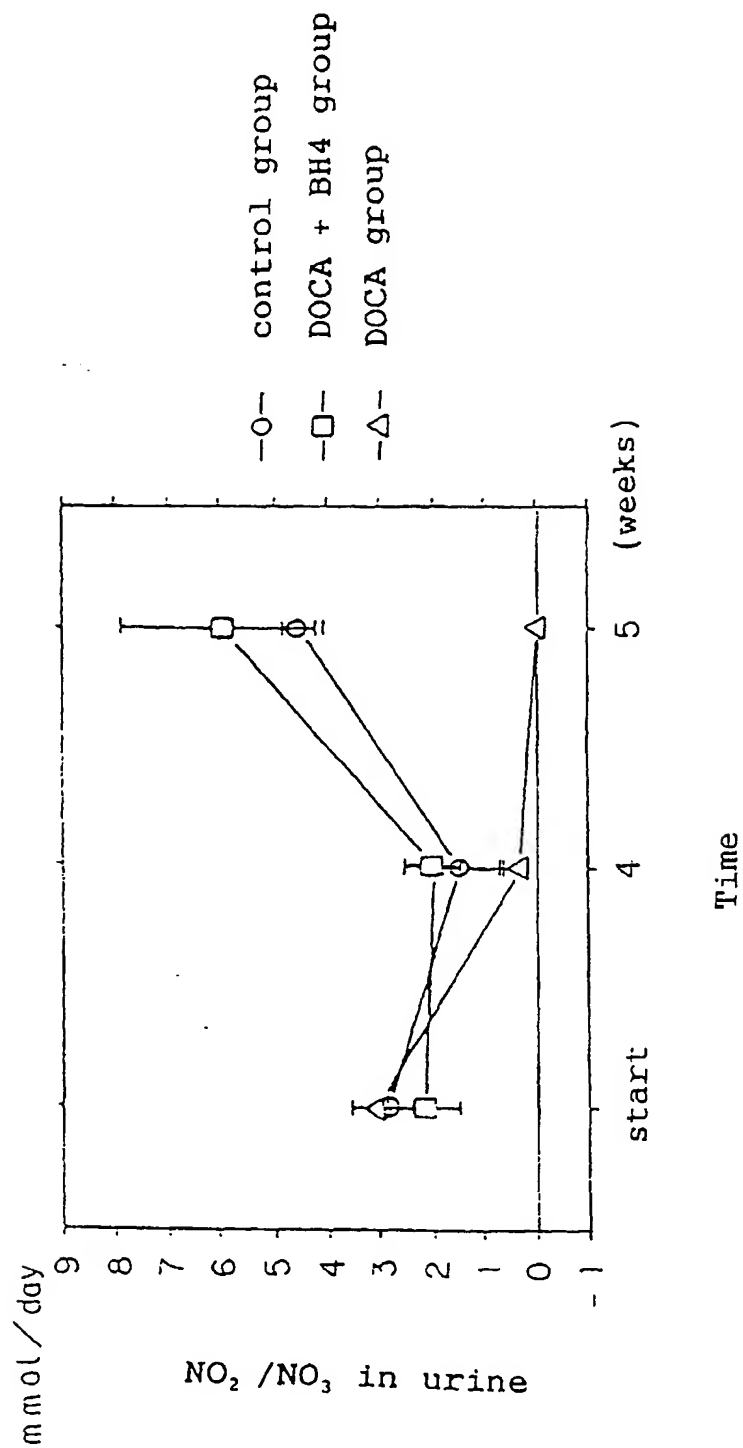


Fig. 3

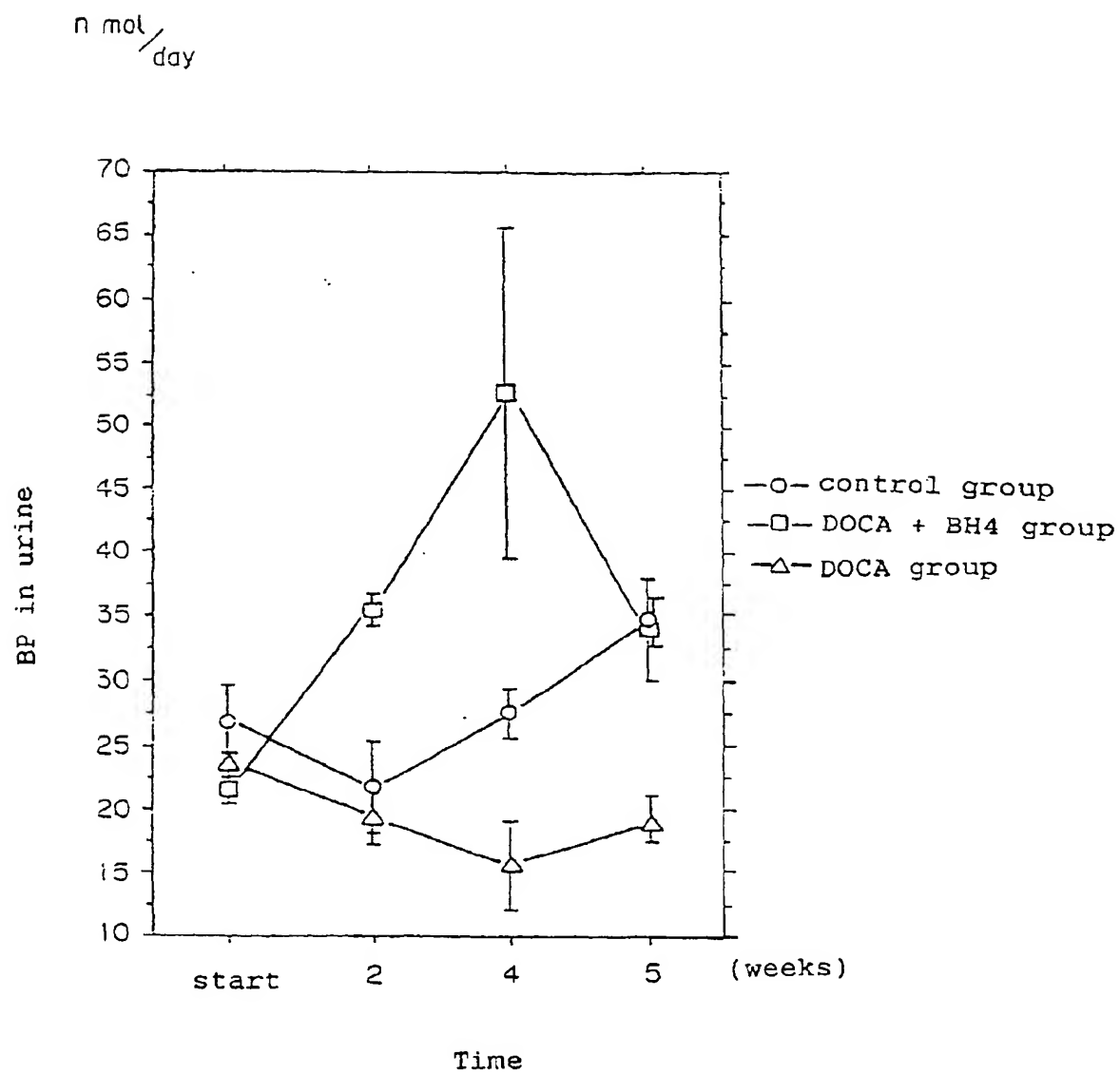


Fig. 4

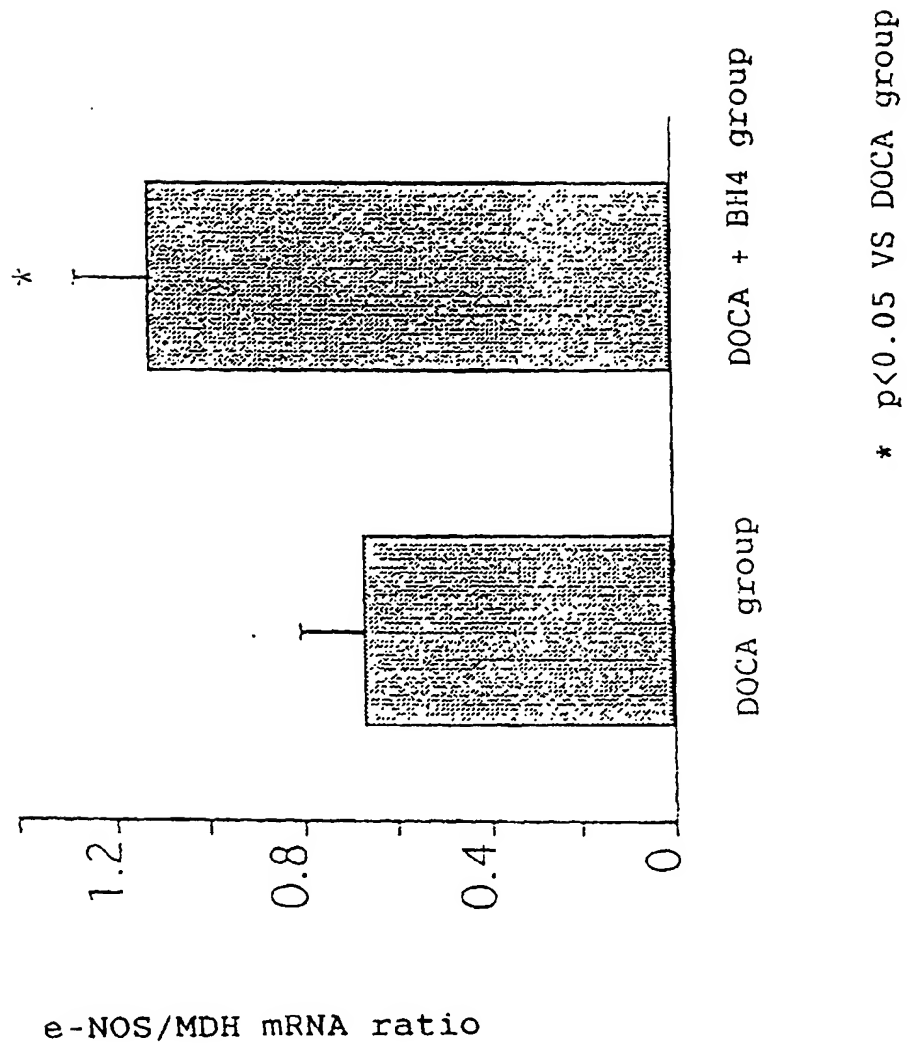


Fig. 5

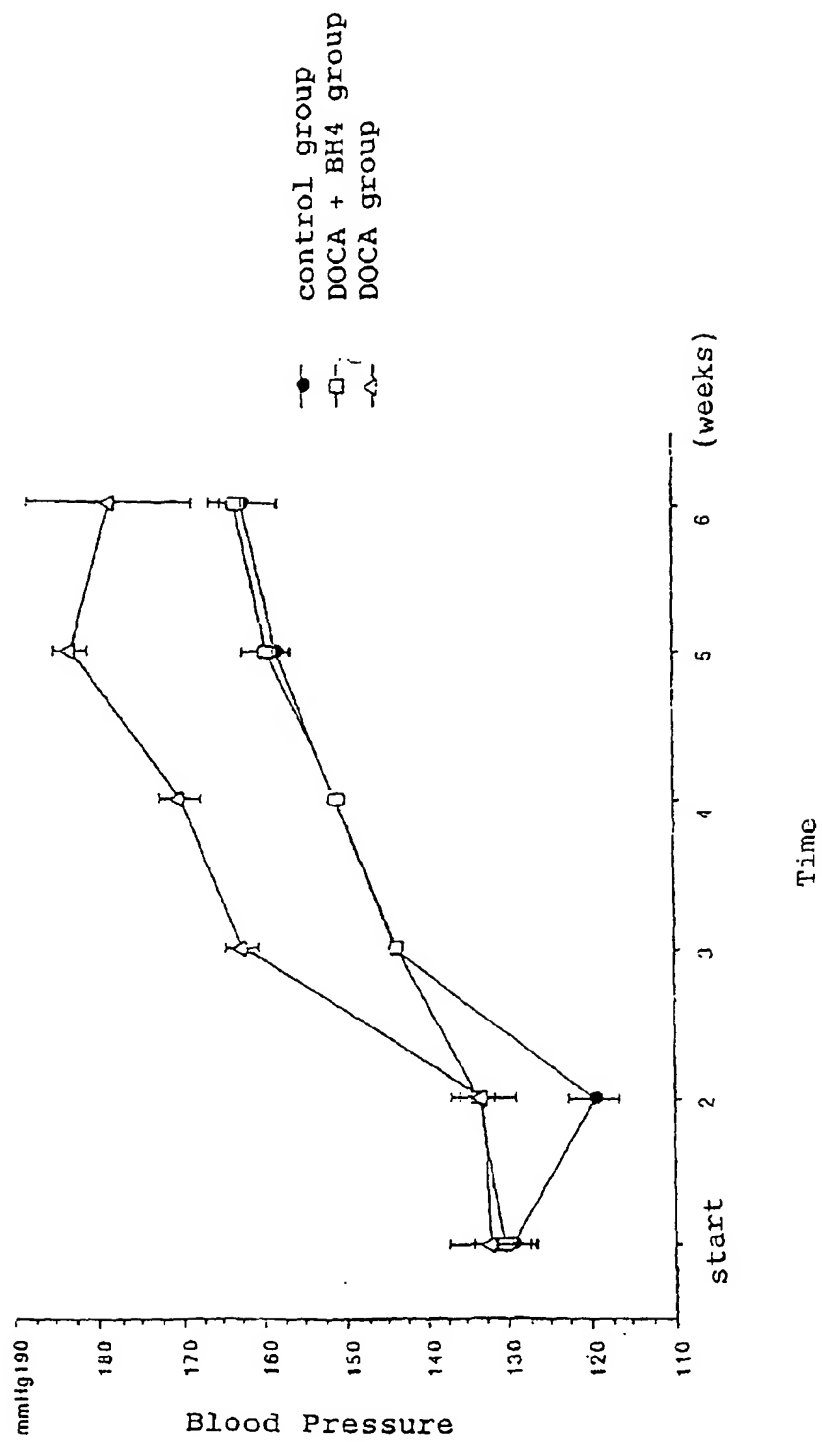


Fig. 6

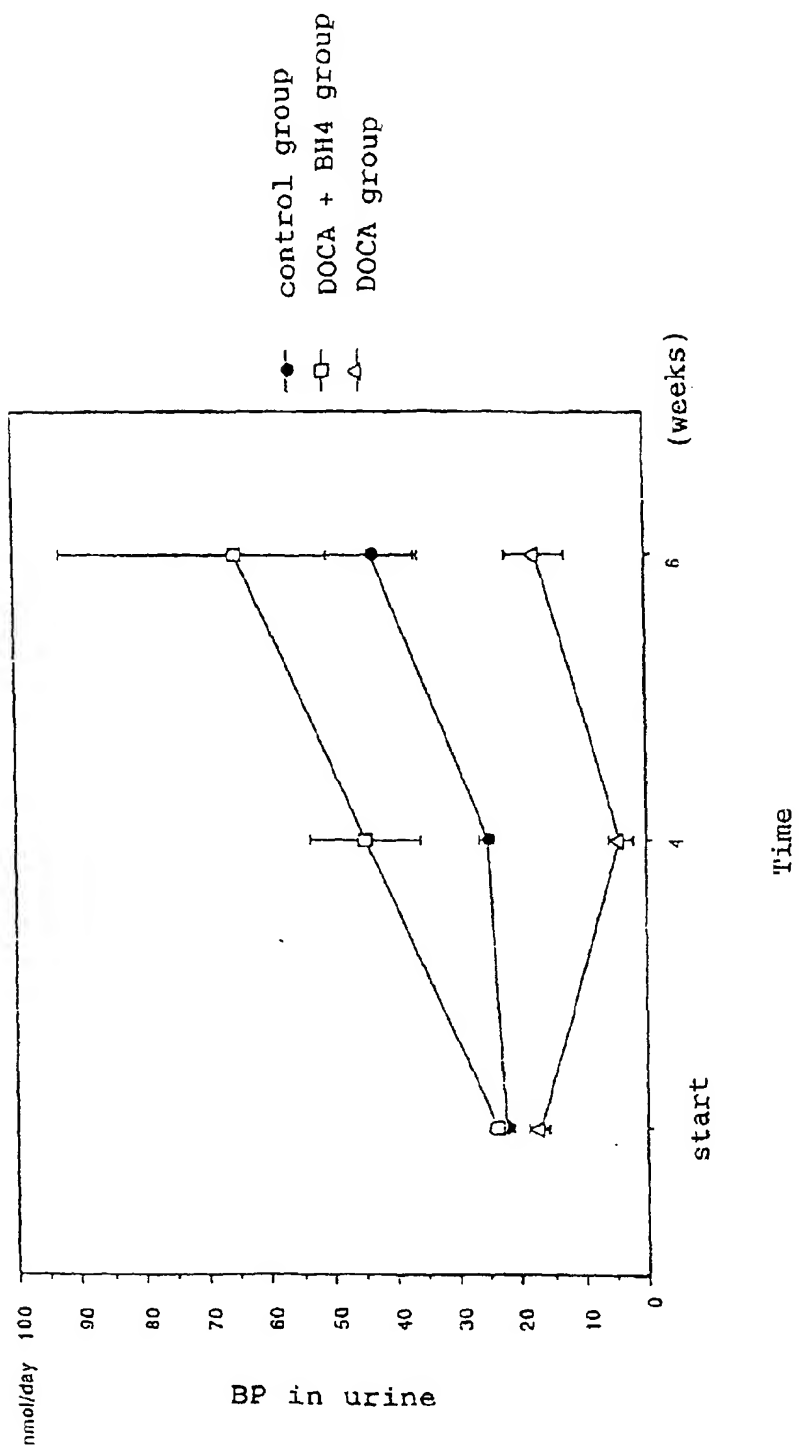


Fig. 7

